

# CLEARTEST® DIAGNOSTIK

## CLEARTEST® Syphilis

A rapid test for the diagnosis of Syphilis to detect antibodies (IgG and IgM) to *Treponema Pallidum* (TP) qualitatively in whole blood, serum or plasma.

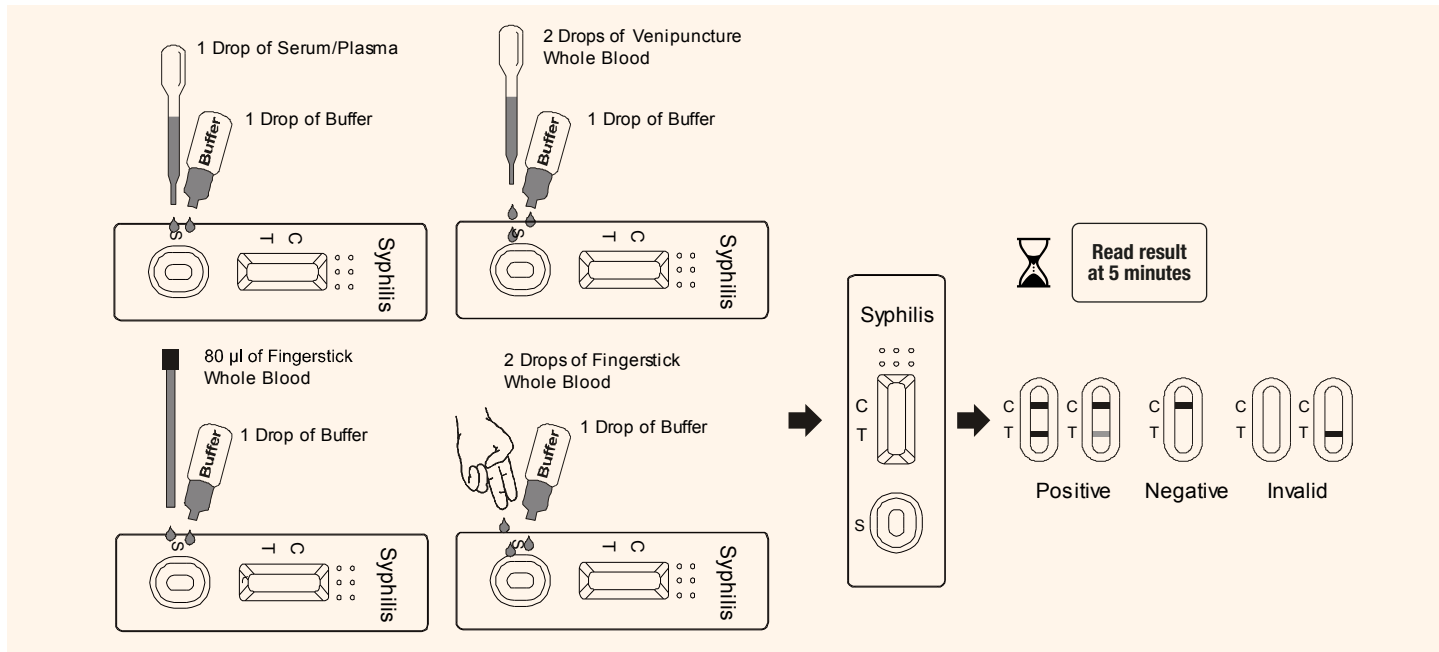
*For professional in vitro diagnostic use only.*

### INSTRUCTIONS FOR USE



#### INTENDED USE

The CLEARTEST® Syphilis is a rapid chromatographic immunoassay for the qualitative detection of antibodies (IgG and IgM) to *Treponema Pallidum* (TP) in whole blood, serum or plasma to aid in the diagnosis of Syphilis.



#### SUMMARY

*Treponema Pallidum* (TP) is the causative agent of the venereal disease Syphilis. TP is a spirochete bacterium with an outer envelope and a cytoplasmic membrane.<sup>1</sup> Relatively little is known about the organism in comparison with other bacterial pathogens. According to the Center for Disease Control (CDC), the number of cases of Syphilis infection has markedly increased since 1985.<sup>2</sup> Some key factors that have contributed to this rise include the crack cocaine epidemic and the high incidence of prostitution among drug users.<sup>3</sup> One study reported a substantial epidemiological correlation between the acquisition and transmission of the HIV virus and Syphilis.<sup>4</sup>

Multiple clinical stages and long periods of latent, asymptomatic infection are characteristic of Syphilis. Primary Syphilis is defined by the presence of a chancre at the site of inoculation. The antibodies response to the TP bacterium can be detected within 4 to 7 days after the chancre appears. The infection remains detectable until the patient receives adequate treatment.<sup>5</sup>

The CLEARTEST® Syphilis utilizes a double antigen combination of a Syphilis antigen coated particle and Syphilis antigen immobilized on membrane to detect TP antibodies (IgG and IgM) qualitatively and selectively in whole blood, serum or plasma.

#### PRINCIPLE

The CLEARTEST® Syphilis is a qualitative membrane based immunoassay for the detection of TP antibodies (IgG and IgM) in whole blood, serum or plasma. In this test procedure, recombinant Syphilis antigen is immobilized in the test line region of the test. After specimen is added to the specimen well of the cassette, it reacts with Syphilis antigen coated particles in the test. This mixture migrates chromatographically along the length of the test

and interacts with the immobilized Syphilis antigen. The double antigen test format can detect both IgG and IgM in specimens. If the specimen contains TP antibodies, a colored line will appear in the test line region, indicating a positive result. If the specimen does not contain TP antibodies, a colored line will not appear in this region, indicating a negative result. To serve as a procedural control, a colored line will always appear in the control line region, indicating that proper volume of specimen has been added and membrane wicking has occurred.

#### REAGENTS

The test contains Syphilis antigen coated particles and Syphilis antigen coated on the membrane.

#### PRECAUTIONS

- For professional in vitro diagnostic use only. Do not use after expiration date.
- Do not eat, drink or smoke in the area where the specimens or kits are handled.
- Do not use test if pouch is damaged.
- Handle all specimens as if they contain infectious agents. Observe established precautions against microbiological hazards throughout all procedures and follow the standard procedures for proper disposal of specimens.
- Wear protective clothing such as laboratory coats, disposable gloves and eye protection when specimens are assayed.
- The used test should be discarded according to local regulations.
- Humidity and temperature can adversely affect results.

## STORAGE AND STABILITY

Store as packaged in the sealed pouch either at room temperature or refrigerated (2–30 °C). The test is stable through the expiration date printed on the sealed pouch. The test must remain in the sealed pouch until use. **DO NOT FREEZE.** Do not use after the expiration date.

## SPECIMEN COLLECTION AND PREPARATION

- The CLEARTEST® Syphilis can be performed using whole blood (from venipuncture or fingerstick), serum or plasma.
- To collect **Fingerstick Whole Blood specimens:**
  - Wash the patient’s hand with soap and warm water or clean with an alcohol swab. Allow to dry.
  - Massage the hand without touching the puncture site by rubbing down the hand towards the fingertip of the middle or ring finger.
  - Puncture the skin with a sterile lancet. Wipe away the first sign of blood.
  - Gently rub the hand from wrist to palm to finger to form a rounded drop of blood over the puncture site.
  - Add the Fingerstick Whole Blood specimen to the test by using **a capillary tube:**
    - Touch the end of the capillary tube to the blood until filled to approximately 80µL. Avoid air bubbles.
    - Place the bulb onto the top end of the capillary tube, then squeeze the bulb to dispense the whole blood to the specimen area of the test cassette.
  - Add the Fingerstick Whole Blood specimen to the test by using **hanging drops:**
    - Position the patient’s finger so that the drop of blood is just above the specimen area of the test cassette.
    - Allow 2 hanging drops of fingerstick whole blood to fall into the center of the specimen area on the test cassette, or move the patient’s finger so that the hanging drop touches the center of the specimen area. Avoid touching the finger directly to the specimen area.
- Separate serum or plasma from blood as soon as possible to avoid hemolysis. Use only clear non-hemolyzed specimens.
- Testing should be performed immediately after the specimens have been collected. Do not leave the specimens at room temperature for prolonged periods. Serum and plasma specimens may be stored at 2–8 °C for up to 3 days. For long term storage, specimens should be kept below –20 °C. Whole blood collected by venipuncture should be stored at 2–8 °C if the test is to be run within 2 days of collection. Do not freeze whole blood specimens. Whole blood collected by fingerstick should be tested immediately.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Specimens should not be frozen and thawed repeatedly.
- If specimens are to be shipped, they should be packed in compliance with local regulations covering the transportation of etiologic agents.

## MATERIALS

### Materials provided

- Test Cassettes
- Droppers
- Buffer
- Instruction for use

### Materials required but not provided

- Specimen collection Containers
- Centrifuge
- Timer
- Lancets
- Heparinized capillary tubes and dispensing bulb

## DIRECTIONS FOR USE

**Allow the test, specimen, buffer and/or controls to reach room temperature (15–30 °C) prior to testing.**

1. Bring the pouch to room temperature before opening it. Remove the test cassette from the sealed pouch and use it as soon as possible.
2. Place the cassette on a clean and level surface.
  - For **Serum or Plasma** specimen: Hold the dropper vertically and **transfer 1 drop of serum or plasma** (approximately 40µL) to the specimen area, then **add 1 drop of buffer** (approximately 40µL), and start the timer, see illustration below.
  - For **Venipuncture Whole Blood** specimen: Hold the dropper vertically and **transfer 2 drops of whole blood** (approximately 80µL) to the specimen area, then **add 1 drop of buffer** (approximately 40µL), and start the timer. See illustration on the first page.
  - For **Fingerstick Whole Blood** specimen:
    - To use a capillary tube: Fill the capillary tube **and transfer approximately 80µL of fingerstick whole blood specimen** to the specimen area of test cassette, then **add 1 drop of buffer** (approximately 40µL) and start the timer. See illustration on the first page.
    - To use hanging drops: Allow **2 hanging drops of fingerstick whole blood specimen** (approximately 80µL) to fall into the specimen area of test cassette, then **add 1 drop of buffer** (approximately 40µL) and start the timer. See illustration on the first page.
3. Wait for the colored line(s) to appear. **Read results at 5 minutes.** Do not interpret the result after 20 minutes.

## INTERPRETATION OF RESULTS

(Please refer to the illustration on the first page)

**POSITIVE:\* Two lines appear.** One colored line should be in the control line region (C) and another apparent colored line should be in the test line region (T).

**\*NOTE:** The intensity of the color in the test line region (T) will vary depending on the concentration of TP antibodies present in the specimen. Therefore, any shade of color in the test line region (T) should be considered positive.

**NEGATIVE: One colored line appears in the control line region (C).** No line appears in the test line region (T).

**INVALID: Control line fails to appear.** Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test. If the problem persists, discontinue using the test kit immediately and contact your local distributor.

## QUALITY CONTROL

A procedural control is included in the test. A colored line appearing in the control line region (C) is considered an internal procedural control. It confirms sufficient specimen volume, adequate membrane wicking and correct procedural technique.

Control standards are not supplied with this kit; however, it is recommended that positive and negative controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance.

## LIMITATIONS

1. The CLEARTEST® Syphilis is for in vitro diagnostic use only. The test should be used for the detection of TP antibodies in whole blood, serum or plasma specimens only. Neither the quantitative value nor the rate of increase in TP antibodies can be determined by this qualitative test.
2. The CLEARTEST® Syphilis will only indicate the presence of TP antibodies in the specimen and should not be used as the sole criteria for the diagnosis of TP infection.
3. As with all diagnostic tests, all results must be interpreted together with other clinical information available to the physician.
4. If the test result is negative and clinical symptoms persist, additional testing using other clinical methods is recommended. A negative result does not at any time preclude the possibility of TP infection.

## EXPECTED VALUES

The CLEARTEST® Syphilis has been compared with a leading commercial TPPA Syphilis test, demonstrating an overall accuracy greater than or equal to 99.8 %.

## PERFORMANCE CHARACTERISTICS

### Sensitivity and Specificity

The CLEARTEST® Syphilis has correctly identified specimens of a performance panel and has been compared to a leading commercial TPPA Syphilis test using clinical specimens. The results show that the relative sensitivity of the CLEARTEST® Syphilis is >99.9 % and the relative specificity is 99.7 %.

Method	Results	TPPA		Total Result
		Positive	Negative	
CLEARTEST® Syphilis	Positive	200	1	201
	Negative	0	319	319
Total Result		200	320	520

Relative Sensitivity: >99.9 % (95 %CI\*: 99.4 % – 100 %)

Relative Specificity: 99.7 % (95 %CI\*: 98.3 % – 100 %)

Accuracy: 99.8 % (95 %CI\*: 98.9 % – 100 %)

\*Confidence Interval

## PRECISION

### Intra-Assay

Within-run precision has been determined by using 10 replicates of four specimens: a negative, a low positive, a medium positive and a high positive. The negative, low positive, medium positive and high positive values were correctly identified >99 % of the time.

### Inter-Assay

Between-run precision has been determined by 10 independent assays on the same four specimens: a negative, a low positive, a medium positive and a high positive. Three different lots of the Syphilis Rapid Test Cassette (Whole Blood/Serum/Plasma) have been tested over a 3-day period using negative, low positive, medium positive and high positive specimens. The specimens were correctly identified >99 % of the time.

### Cross-reactivity

The CLEARTEST® Syphilis has been tested by HAMA, RF, HBsAg, HBsAb, HBeAg, HBeAb, HBcAb, HCV, HIV, H. Pylori, MONO, CMV, Rubella and TOXO positive specimens. The results showed no cross-reactivity.

## Interfering Substances

The following potentially interfering substances were added to Syphilis negative and positive specimens.
















Acetaminophen: 20 mg/dL	Caffeine: 20 mg/dL
Acetylsalicylic Acid: 20 mg/dL	Gentisic Acid: 20 mg/dL
Ascorbic Acid: 2 g/dL	Albumin: 2 g/dL
Creatin: 200 mg/dL	Hemoglobin 1.1 mg/dL
Bilirubin: 1 g/dL	Oxalic Acid: 600 mg/dL

None of the substances at the concentration tested interfered in the assay.

## BIBLIOGRAPHY

1. Claire M. Fraser. Complete genome sequence of *Treponema Pallidum*, the Syphilis spirochete, Science 1998; 281 July: 375-381
2. Center for Disease Control. Recommendations for diagnosing and treating Syphilis in HIV-infected patients, MMWR Morb. Mortal Wkly Rep. 1988; 37: 601
3. Aral R. Marx. Crack, sex and STD, Sexually Transmitted Diseases, 1991; 18:92-101
4. J.N. Wasserheit. Epidemiological Synergy: Interrelationships between human immunodeficiency virus infection and other sexually transmitted diseases, Sexually Transmitted Diseases 1992; 19:61-77
5. Johnson Phillip C. Testing for Syphilis, Dermatologic Clinic 1994; 12 Jan: 9-17

## Index of Symbols

Index of symbols			
	Article number		Temperature limitation
	Observe operating instructions		Batch number
	In-vitro-diagnostic		Expiry date
	Manufacturer		Content sufficient for <n> tests
	Dangerous substances		Single use
	Protect from heat and sunlight		Attention
	Protect from moisture		
	Do not use, if package is damaged		
	CE marked according to IVD directive 98/79/EG		

## ORDERING INFORMATION

### CLEARTEST® Syphilis 5-test cassettes

 C3 0610-05

 00524134

### CLEARTEST® Syphilis 10-test cassettes


 C3 0610-10

 0052410



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